REVIEW

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The cGAS-STING pathway in COPD: targeting its role and therapeutic potential



Kexin Liao¹, Fengshuo Wang³, Chenhao Xia², Ze Xu², Sen Zhong², Wenqi Bi¹ and Jingjing Ruan^{2*}

Abstract

Chronic obstructive pulmonary disease(COPD) is a gradually worsening and fatal heterogeneous lung disease characterized by airflow limitation and increasingly decline in lung function. Currently, it is one of the leading causes of death worldwide. The consistent feature of COPD is airway inflammation. Several inflammatory factors are known to be involved in COPD pathogenesis; however, anti-inflammatory therapy is not the first-line treatment for COPD. Although bronchodilators, corticosteroids and roflumilast could improve airflow and control symptoms, they could not reverse the disease. The cyclic GMP-AMP synthase-stimulator of interferon genes (cGAS-STING) signaling pathway plays an important novel role in the immune system and has been confirmed to be a key mediator of inflammation during infection, cellular stress, and tissue damage. Recent studies have emphasized that abnormal activation of cGAS-STING contributes to COPD, providing a direction for new treatments that we urgently need to develop. Here, we focused on the cGAS-STING pathway, providing insight into its molecular mechanism and summarizing the current knowledge on the role of the cGAS-STING pathway in COPD. Moreover, we explored antagonists of cGAS and STING to identify potential therapeutic strategies for COPD that target the cGAS-STING pathway.

Keywords cGAS-STING pathway, COPD, Agonists, Therapeutic potential

Introduction

COPD is a progressive and debilitating respiratory disease that affects millions of people worldwide and poses a considerable medical and financial burden [1, 2]. Traditionally, COPD is considered as an inflammatory response elicited by cigarette smoking(CS) in older males [3]. In addition, other factors, such as air pollution, occupational particles and aging, have also been found

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to trigger lung inflammation, with COPD subsequently accompanied by inflammation [2].

Pattern recognition receptors (PRRs) are a significant component of the innate immune system and constitute the first line of defense in organisms. As a member of the PPR family, the cGAS protein acts as an innate nucleic acid sensor recognizing exogenous DNA generated by viral or bacterial infection or in the cytoplasm and converts ATP and GTP into 2'3'-cyclic GMP-AMP (cGAMP), which can be used to monitor pathogen infection or cellular stress [4]. cGAMP binds to the adapter protein stimulator of interferon genes (STING) localized at the endoplasmic reticulum (ER) membrane [5] and initiates a downstream immune response. Increasing research suggests that the cGAS-STING pathway plays an important role in the development of many diseases



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through its involvement in autoimmunity, cellular senescence and anti-inflammation [6].

The Global Initiative of Obstructive Lung Disease (GOLD) has suggested guidelines for COPD management. However, symptomatic treatment involving bronchodilators continues to be the mainstay in COPD management, despite the understanding of inflammation as a key driver of COPD progression. There is currently no cure for COPD. Recent efforts have tended to focus on the molecular mechanisms underlying COPD to explore therapeutic targets for COPD. Studies have shown that cGAS-STING contributes to COPD, especially under exposure to cigarette smoking [7] or air pollutants including silica [8] and PM 2.5 [9]. Moreover, targeting the cGAS-STING pathway can circumvent cellular senescence [10, 11], which has also been shown to contribute to the accelerated aging process in COPD patients [12]. Delving into the structure and function of the cGAS-STING pathway may enable the development of selective small-molecule inhibitors to manage the inflammation associated with COPD. In this review, we discussed the role of the cGAS-STING pathway in the pathogenesis of COPD, as well as antagonists of this pathway, with a focus on its therapeutic potential for COPD. Our aim is to contribute to the optimization of fundamental therapies for COPD, ultimately improving patient prognosis.

Inflammation-associated mechanisms in the pathogenesis of COPD

Airway inflammation is a consistent feature of COPD and plays an important role in the disease pathogenesis, progression and mortality [2, 13]. Inflammation has many manifestations. In this paragraph, we describe neutrophil-associated and eosinophil-associated inflammation in COPD as well as some relevant inflammatory signaling pathways.

Neutrophil-associated airway inflammation in COPD

Neutrophil inflammation is the key inflammatory phenotype in the pathogenesis of COPD, with increased neutrophils in sputum and blood being a characteristic feature of all COPD patients. Studies have reported that neutrophil count is a marker of COPD severity and patients with higher sputum neutrophil percentages have greater dyspnea scores [2, 14, 15]. When stimulated by inflammation, neutrophils leave the circulation to congregate in lungs. The aggregation of neutrophils produces a large amount of reactive oxygen species (ROS), which can destroy lung tissues [16]. Moreover, neutrophils produce the inflammatory factor IL-6, which induces the production of elastase and oxygen free radicals, thereby increasing pulmonary vascular permeability and exacerbating lung tissue destruction [17]. Neutrophils accumulate in the airways of COPD patients [18] and can secrete serine proteases including matrix metalloproteinase (MMP) and neutrophil elastase (NE) [19]. MMP is significantly increased in patients with COPD and destroys the structural components of extracellular matrix (ECM), contributing to alveolar destruction [20]. In animal models, dominant-negative MafB was shown to downregulated MMP, thereby suppressing porcine pancreatic elastase-induced emphysema [21]. NE is a neutrophilderived serine proteinase and has proven to be involved in lung damage. A study revealed that NE deficiency in mice protects them from emphysema after exposure to cigarette smoke (CS) [22]. The underlying mechanism may be that NE can also degrade the structural components of ECM and cooperate with MMPs to amplify the degradation [23]. In addition, NE is effective in stimulating mucus secretion from submucosal glands and thrush cells, leading to airway obstruction [24]. All these findings indicate the contribution of neutrophil inflammation to the development of COPD.

Eosinophilic-associated airway inflammation in COPD

Although neutrophil-associated COPD is the most common inflammatory phenotype, it has been recognized that eosinophils may also be involved in the inflammatory response in COPD. Approximately 10-40% of COPD patients demonstrate increased eosinophilic inflammation in the sputum or blood [25]. Eosinophilic airway inflammation occurs in COPD exacerbations. Clinical research has shown that patients with high eosinophil count but a low percentage of macrophages exhibit the greatest decline in lung function during an exacerbation and a greater exacerbation frequency. This group of patients has persistent eosinophilic inflammation due to defective macrophage efferocytosis, which contributes to the severity of the disease [26]. Like in asthma, recruitment of eosinophils to the airway in COPD is mediated via CCR3 chemokines, which play a critical role together with other eosinophil chemoattractants, such as prostaglandin (PG)D2 [27, 28]. Inflammatory cues prompt the recruitment of eosinophils into the lungs, where the secretion of a variety of chemokines (e.g., CCL5, CCL11, CCL13), cytokines (e.g., IL-2, IL-3, IL-4, IL-5, IL-10, IL-12, IL-13, IL-16, IL-25) and cytotoxic granular products (major basic protein, eosinophil cationic protein, eosinophil peroxidase, eosinophil-derived neurotoxin) contributes to inflammation [29]. An increase in eosinophilic inflammation in peripheral blood and sputum samples from COPD patients is associated with an increased risk of severe deterioration in the future [30]. However, the etiology of eosinophilic inflammation in COPD is not completely understood.

Inflammation-associated pathways in COPD

The pathogenesis of COPD involves the activation of diverse inflammatory pathways. The NF- κ B pathway is activated by the ubiquitination of I κ B [31]. As a result, NF- κ B is released from the NF- κ B/I κ B complex and are able to bind to target genes, thereby initiating the expression of target genes, such as TNF- α and IL-1, and causing an inflammatory response [32]. In addition, the inhalation of ozone and cigarette smoke results in the migration of neutrophils into the lungs to generate ROS, which is another factor in the activation of NF- κ B [33]. A study in mouse models of COPD demonstrated that NF-kB pathway is essential for inflammation in smoking-induced bronchiolitis [34]. Moreover, hypomethylation of NF- κ B-mediated pathway genes has also been conformed to contribute to COPD exacerbation [35].

The mitogen-activated protein kinase (MAPK) pathway participates in stress adaptation and inflammatory responses and its activation can stimulate cytokines, neurotransmitters, serine proteases, and oxidative stress [36]. Haemophilus influenzae is a common pathogen of COPD, and it was found to upregulate MUC gene transcription through the activation of MAPK signaling pathway [37]. In addition, IL-8 and TNF- α are key factors in COPD development and are are also regulated by p38MAPK [38].

Many other pathways including EGFR signaling pathway [39], MARCKS protein signaling pathway [40], SNARE protein signaling pathway [41], and Ciliophagy signaling pathway [42] are associated with airway mucus hypersecretion, which is recognized as one of the main pathophysiological changes in COPD patients. The cGAS-STING signaling pathway is also highly involved, and we elaborate on this pathway in this review.

Overview of the cGAS-STING pathway

Sun et al. identified cGAS through isolation and purification in 2013 [43], and revealed a novel immune signaling pathway, namely, the cGAS-STING signaling pathway. This pathway occurs within cells and is highly important for immune systems that can sense double-stranded DNA (dsDNA) to defend against extracellular or intracellular pathogens [4]. The cGAS-STING pathway has emerged as a critical mechanism for the induction of powerful innate immune defense programs [44].

In many organisms, the detection of foreign DNA is a key factor in immunity. In mammalian cells, this process is largely facilitated by the cGAS, which has become an important mechanism for combining DNA perception with the induction of powerful innate immune defense strategies [45, 46]. PRRs are essential components of the innate immune system that can recognize biomolecules such as pathogen-associated molecular patterns (PAMPs) and DAMPs. PAMPs contain double- or single-stranded

DNA and RNA generated by viral or bacterial infection or in the cytoplasm [47]. The strongest response after DNA stimulation is initiated by cGAS, a member of the PRR family, which is activated after binding to dsDNA [43] in a minimal 2:2 complex to induce conformational changes that allow cGAS to catalyze ATP and GTP into 2,3'-cGAMP [48-50]. The sugar phosphate backbone of DNA-binds to a nucleotide transferase domain (catalytic part) in the C-terminus of cGAS, which includes positively charged DNA binding sites, a primary site, and two additional sites [45]. cGAMP binds to STING, inducing its phosphorylation of STING and causing its conformational changes, activating downstream signal transduction. During this process, STING undergoes high-order oligomerization to form tetramers [51, 52] and is transferred from the endoplasmic reticulum to the intermediate compartment of the endoplasmic reticulum Golgi apparatus. For the past several years, structural studies have shown that the tetramerization of STING in the Golgi complex is a signaling platform for recruiting and activating dimeric TANK-binding kinase 1 (TBK1) dimers through phosphorylation [53]. Conversely, TBK1 transphosphorylates the C-terminal domain of STING to recruit interferon regulatory factor 3 (IRF3) for activation [54], where IRF3 translocates to the nucleus. This gene plays a transcriptional role in the expression of immune stimulating genes (ISGs) and type 1 interferons (IFNs) [43, 50]. Moreover, STING also activated IKB kinase (IKK)-mediated induction of NF-kB-driven inflammatory genes. After activation, STING is transported to the inner lysosome for degradation, while NF-KB translocates to the nucleus, where it triggers the the expression of proinflammation cytokines(e.g., TNF and IL-6) [55, 56]. Activated STING passes through signal transduction pathways, ultimately leading to the production of a large amount of interferon and other immune related cytokines, thereby triggering an immune response. In addition, the binding of cGAS to DNA is independent of the DNA sequence [57]. Therefore, theoretically, self DNA from mitochondria or nuclei can also act as a cGAS ligand, activating the cGAS-STING pathway and triggering inflammatory responses [58]. Recent studies have also shown that endogenous cGAS is tightly bound to the nucleus and prevents its self response to its own DNA [59, 60]. Moreover, other studies have shown that cGAS inhibits homologous recombination mediated DNA repair and promotes genomic instability, micronucleus generation, and cell death under genomic stress conditions in a manner independent of STING by other studies [61]. With increasing researches on cGAS and STING, the cGAS-STING pathway has been revealed to be involved in autoimmunity, cellular senescence and inflammation inhibition, indicating that it plays an

important role in the occurrence of inflammation and many diseases [6, 62, 63].(Fig. 1).

A schematic detailing the cGAS-STING signaling pathway. Upon binding dsDNA, cGAS dimers assemble on dsDNA to generate 2'-3'cGAMP. cGAMP binds to STING, leading to the translocation of STING from the ER to the Golgi and ER-Golgi intermediate compartment (ERGIC). This activation of STING recruits TBK1 and IKK, promoting their autophosphorylation and triggering the phosphorylation of IRF3 and IkB. Phosphorylated IRF3 translocates to the nucleus, where it results in the gene expression of type I interferons. Phosphorylated IkB recruits NF-kB, induces expression of genes encoding proinflammatory cytokines, and is subsequently degraded.

The cGAS-STING pathway in COPD

The cGAS-STING pathway in cigarette smoke-induced COPD

The significance of the cGAS-STING pathway in the inflammatory response is well known. As mentioned

earlier, smoking is a key factor that induces COPD. In 2016, Pouwels revealed that exposure to a smoke environment can induce the release of dsDNA and mtDNA in mouse bronchoalveolar tissues in vivo, leading to cell death in human bronchial epithelial cells, and increased release of dsDNA and mtDNA was detected in the extracellular environment [64]. Moreover, sensing of these two types of DNA opens up the cGAS-STING immune pathway. In a 2019 study, Sears reported that exposure to CS triggers cGAS and STING expression at the mRNA and protein levels. Importantly, DNA damage repair defects are related to the pathogenesis of COPD, indicating that DNA release and sensing play a crucial role [65]. In mice and COPD patients, macrophage uptake of nanoparticulate carbon black induces DNA repair enzymes, leading to dsDNA breakage and an inflammatory response through activation of the cGAS-STING signaling pathway [66]. Moreover, in 2019, Nascimento, using genedeficient mouse strains, reported that the absence of cGAS or STING can lead to reduced lung inflammation. In this study, the BALF of COPD model mice showed



Fig. 1 Overview of the cGAS-STING pathway

overexpression of cGAS and STING, while the DNA content increased, and neutrophil recruitment increased. Compared with wild-type mice, mice with STING and cGAS gene knockout showed a significant decrease in dsDNA content in the BALF and in the production of the downstream chemokine CXCL10 and relatively mild lung inflammation, while TLR-9 gene knockout mice showed no significant changes. This finding suggested that the self-DNA released after exposure to cigarette smoke is recognized by cGAS rather than by TLR-9, which activates the STING pathway and leads to increased secretion of type I IFN, promoting pulmonary inflammation [7]. In addition, the release of some self-dsDNA is dependent on STING, indicating that lung injury induces de novo cell death and self-dsDNA-dependent lung inflammation through amplification loops [67].

The cGAS-STING pathway in particulate matter-induced COPD

The long-term inhalation of particulate matter, such as silica and PM 2.5, is another major cause of COPD and can also lead to chronic lung inflammation. Exposure to silica particles can induce the release of proinflammatory and profibrotic factors (e.g., IL-6, TNF- α , and TGF- β), which contribute to the acceleration of lung inflammation, and the activation of the cGAS-STING signaling pathway is involved in this process. Benmerzoug reported that mitochondria can be a source of self-dsDNA triggering DNA sensor activation after exposure to silica particles, triggering the type I IFN pathway and inducing cell death in the lungs [8]. After silica exposure, both the

Table 1	cGAS and	STING	antac	onists
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cGAS and STING Antagonists	Name	Molecular mechanism	Ref- er-
			ences
cGAS inhibitors	Hydroxychloroquine	Disrupting DNA binding	[69]
	Quinacrine		[69]
	Хб		[70]
	A151		[71]
	Suramin		[72]
	RU.521	Occupying cGAS Catalytic site	[73]
	PF-06928125		[74]
STING inhibitors	Tetrahydroisoquinoli- none acetate (Com- pound 18)	Targeting the CDN-binding site	[75]
	Astin C		[76]
	Nitrofuran derivatives(C-170, C-171, etc.)	Targeting STING palmitoylation sites	[77]
Unknown inhibitors	CU-32 and CU-76	unknown mechanisms	[78]
	VS-X4		[79]

STING and NLRP3 pathways were activated, leading to cell death and the release of proinflammatory cytokines. This process leads to necrosis and apoptosis in a STING-dependent manner. Another study conducted by Wang in 2022 [9] revealed that PM2.5-induced aging is regulated by an inflammatory response that is activated by the cGAS/STING/NF- κ B pathway, which is closely related to DNA damage. Their study also showed that pretreatment with selenomethionine (Se Met) can inhibit the inflammatory response and prevent cell aging by blocking the cGAS/STING pathway in A549 cells exposed to PM2.5. In addition, the in vivo C57BL/6J mouse model showed a decrease in cGAS expression after Se Met treatment, which can alleviate PM2.5-induced lung tissue aging in mice.

Collectively, the studies discussed here have established the implications and characteristics of STING signaling activation in COPD development. These findings suggest that known COPD-causative factors (e.g., CS, silica, and PM 2.5) can trigger the activation of the cGAS-STING signaling pathway and that targeting this pathway could help alleviate inflammation in COPD patients. Nonetheless, most of the studies discussed here were mouse model-based; therefore, there is an urgent need for human-based research to elucidate the involvement of STING in COPD.

Antagonists of the cGAS-STING pathway

There is no cure for COPD; however, the emergence of the first drug-like compounds selectively targeting cGAS or STING has opened the door for the development of clinical candidates. Several small-molecule agonists have been developed and are being tested in tumor immunotherapy [68]. Although the cGAS-STING pathway antagonists have the optimal beneficial effects on tumor diseases, prominent efforts are underway to develop novel compounds to control the severe inflammation and acute tissue damage observed from chronic stimulation by antagonizing cGAS and STING. (Table 1)

Inhibitors of cGAS

Numerous cGAS antagonists have been identified as favorable targets for ameliorating cGAS- and STINGdependent inflammatory diseases. One example is antimalarial drugs (e.g., hydroxychloroquine, quinacrine, suramin, and oligodeoxynucleotides A151 and X6), which specifically bind to two drug sites on the 2:2 cGAS/ dsDNA dimer and have the potential to suppress cGAS activity [69, 71, 72]. At each site, the antimalarial drug acts in the dsDNA minor groove between the cGAS/ DNA interface (the interface connecting the dsDNAbinding site A/B on two neighboring monomers). This results in the interaction of the antimalarial drug with the DNA-binding sites (A and B) on the two neighboring cGAS monomers, which alters the stability of the cGAS/ dsDNA complex and thus inhibits the activation of cGAS by dsDNA and its enzymatic activities. Follow-up studies have indicated that in the presence of the antimalarial drugs studied, IFN- β expression, cGAMP production, and the levels of a number of dsDNA-stimulated cytokines (IL-6 and TNF- α) are inhibited.

Furthermore, RU.521 affected cGAS activity by occupying the catalytic site of cGAS and decreasing its binding affinity for ATP and GTP without directly interfering with dsDNA binding, as identified in mouse studies [73]. It was demonstrated to reduce Ifnb1 mRNA expression in bone marrow-derived macrophages from Trex1-/mice. PF-06928125 has also been shown to act as a cGAS inhibitor [74]. High-throughput biochemical screening of cGAS inhibitors identified PF-06928215. Although PF-06928215 was also able to bind to the cGAS active site with a high affinity value of 0.2 µmol/L and inhibit cGAS activity with an IC50 value of 4.9 µmol/L, PF-06928215 showed no activity in the cellular cGAS assay [80].

Inhibitors of STING

STING is the critical signaling molecule for the cGAS-STING pathway; therefore, developing antagonists of STING may exploit cGAS-STING inhibitors. Haag et al. reported that nitrofuran derivatives, including C-170, C-171, C-176, C-178, and H-151, can block the STINGmediated signaling pathway by covalently modifying the Cys91 residue of STING [77]. Cys91 in STING has been shown to be targeted by the covalent ligand BPK-25, which inhibits STING activation by disrupting the binding of the cyclic dinucleotide ligand cGAMP [81]. Moreover, Tetrahydroisoquinolinone acetate (Compound 18) stabilizes the open, inactive conformation of STING and binds to the cGAMP binding site in a 2:1 ratio, displacing cGAMP from its binding site on STING. Compound 18 potently inhibited in vitro cGAMP-dependent signaling and displayed slow dissociation kinetics and good oral bioavailability [75]. Astin C is a natural cyclopeptide derived from the traditional Chinese medicinal plant Aster tataricus and was identified by Li et al. as a potent bioactive compound that restricts the cGAS-STING signaling to alleviate autoinflammatory response in a Trex1-/- mouse model and in macrophages [76]. It was demonstrated that astin C binds competitively to the CDN site via pull-down experiments using biotinylated astin C and human STING. Astin C blocks the recruitment of IRF3 to the STING signalosome, thus preventing downstream signaling through this pathway.

Inhibitors of unknown mechanisms

Aside from the findings discussed above, there are also several inhibitors whose mechanisms are still unknown. For example, the small molecules CU-32 and CU-76 bind to cGAS without disrupting the binding between cGAS and dsDNA; these small molecules can inhibit the protein-protein interactions (PPIs), interfaces required for IRF3 activation and downstream IFN-I induction, in human monocyte THP-1 cells, but the exact mechanism is still unclear [78]. Additionally, the small heterocyclic compound VS-X4 has been shown to inhibit STING with no elucidated mechanism of action [79]. Further research is needed to reveal the specific mechanisms of action of these inhibitors.

cGAS-STING inhibitors in clinical trials

Currently, some of these compounds are being phased into clinical studies. María Gómez Antúnez reported a higher survival rate in COPD patients hospitalized with SARS-CoV-2 treated with hydroxychloroquine, but they only recommend its use in clinical trials [82]. Moreover, studies has shown that Quinacrine is a potential treatment for COVID-19 virus infection [83] and that Nitrofuran is a therapy for uncomplicated lower urinary tract infection in women [84], indicating their antiinflammatory roles. However, many of these studied inhibitors primarily treat tumors rather than COPD or other inflammatory diseases. More researches should be devoted to controlling inflammation through antagonizing the cGAS-STING pathway, and we expect that more inhibitors of cGAS-STING will enter clinical trials for COPD treatment in the future.

Targeting the cGAS-STING signaling pathway alleviates COPD

A better understanding of the cGAS-STING signaling pathway has led to the identification of several potential therapies for inhibiting inflammation; these therapies have been termed "protectors of COPD patients". Recent studies focused on cGAS and STING have provided new directions for treating COPD.

Circumventing cellular senescence attenuates COPD by targeting the cGAS-STING signaling pathway

Cellular senescence is a state of cell cycle arrest and is among the 9 hallmarks of aging proposed by López-Otín in a landmark paper [85]. Senescent cells including alveolar epithelial and endothelial cells accumulate in the lungs COPD patients [86, 87], and CS-induced oxidative stress is likely to play an important role in the induction of senescence in COPD [88]. In a mouse model-based study, Takao demonstrated the induction of senescence in lung parenchymal cells during the progression of COPD [89]. And evidence from clinical samples of primary human bronchial epithelial cells and lung homogenates from COPD patients indicates the same conclusion [90].

Studies have shown that interfering with DNA binding to the DNA sensor cGAS can induce cellular senescence [10, 11]. Aging-accelerated factors induce the proinflammatory Senescence Associated Secretory Phenotype (SASP), leading to leakage of DNA into the cytoplasm and triggering of the cGAS-STING pathway of the innate immune response [12]. The cGAS-STING also induces the SASP phenotype by accumulating cytoplasmic DNA during senescence [10, 91], thus aggravating the aging response. Wang et al. reported that Se-Met treatment prevents PM2.5-induced senescence via attenuating inflammatory response regulated by cGAS/STING/ NF- κ B pathway, and further causes a reduction in COPD [9]. Since cGAS-STING pathway plays important roles in COPD, these studies further indicate that targeting this pathway may circumvent cellular senescence and thus has therapeutic potential for mitigating COPD.

Natural products relieve COPD by targeting the cGAS-STING signaling pathway

The Tanreqing (TRQ) injection is a Chinese patent medicine. It can significantly improve the partial pressure of oxygen (PaO2), partial pressure of carbon dioxide (PaCO2) and lung function in patients with COPD combined with respiratory failure, and is commonly used to treat AECOPD. Several in vivo experimental studies have also revealed that TRQI can reduce the expression of IL-8, TNF- α and mucin 5AC (MUC5A) in alveolar lavage fluid in CS-and LPS-induced rats with COPD, thereby improving the inflammatory response of airway mucosa and inhibiting airway mucus hypersecretion in rats [92]. TRQ injection inhibited STING levels, suggesting that TRQ has therapeutic efficacy by blocking the increase in the cGAS-STING pathway in COPD patients [93]. However, the specific mechanism of action of TRQI for the treatment of COPD is still unclear. With respect to the five traditional Chinese medicines, further experiments are needed to identify the specific components of TRQ that regulate the cGAS-STING pathway and alleviate COPD.

Panax ginseng C.A Meyer (ginseng) root is another important traditional Chinese medicinal herb. Its pharmacologically active constituents have been identified, most notably ginsenosides, which are triterpenoid saponins that include protopanaxadiol (PPD) and protopanaxatriol (PPT) [94]. The PPD group contains ginsenosides Rb1, Rb2, Rb3, Rc, Rd, Rg3 and Rh2 and compound K, while the PPT group comprises ginsenosides Re, Rf, Rg1, Rg2 and Rh1. These compounds possess pharmacological effects, such as antiviral, antioxidant, and immunomodulatory activities, and have potentially relevant effects on COPD, including the inhibition of proinflammatory mediators and cytokines [95]. Recently, studies have shown that ginsenosides can alleviate COPD and reduce lung injury [96, 97]. In a CS-induced BALV/c mouse model, X. Guan et al. reported that ginsenosides Rg3 and Rb3 could negatively regulate PI3K activation, NF-κB activity, and proinflammatory cytokines in a CS-induced BALV/c rat model, basal cells, and a coculture model of bronchial epithelial cells and neutrophils, thus reducing neutrophil migration. Moreover, ginsenosides basically inhibit various COPD-related pathogenesis processes, such as inflammatory responses (TNF- α , IL-6, IL-1 β , NF- κ B induction and translocation), kinase phosphorylation (MAPK and ERK1/2), and oxidative stress (ROS) [98]. Mechanistically, PPD suppressed the cGAS-SING pathway through the activation of AMPK and the inhibition of TNF, IL and NF- κ B. The therapeutic effect of PPD in COPD patients awaits further clinical investigation.

Both Radix Pseudostellariae and Juglanin, which are types of tonic Chinese medicine, have been shown to reduce lung inflammation by inhibiting the cGAS-STING pathway and therefore alleviating COPD [99, 100]. The findings above suggest that natural products, especially Chinese medical herbs, can alleviate inflammation by inhibiting the cGAS-SING pathway, thus exerting therapeutic effects on COPD. This finding provides potential avenues for future drug development and therapeutic strategies for this disease.

Discussion: summary, outstanding questions, and future directions

Airway inflammation is a consistent characteristic of COPD and is related to its pathogenesis and progression. As an innate immune pathway, cGAS-STING plays an undeniable role in immune-related diseases. The cGAS-STING pathway has been shown to alleviate inflammatory responses and lung function damage in COPD patients, indicating its potential as a therapeutic target [7]. This pathway offers a specific and selective means to modulate immune responses, particularly in DNA-induced inflammation associated with COPD. Current preclinical development efforts are focused on several cGAS and STING inhibitors, including antimalarial drugs [77], RU.521 [73], PF-06928215 [74], nitrofuran [77], compound 18 [75] and astin C [76], ect., which are expected to open up new avenues for treating COPD. Furthermore, targeting the cGAS-STING signaling pathway circumvents cellular senescence to attenuate COPD [10, 11] and an increasing number of natural products, mainly Chinese herbs, have been discovered to alleviate COPD via a pharmacological mechanism involving cGAS-STING [93, 98–100].

However, the pathogenesis of COPD is not fully understood. Most relevant studies are animal-based experiments rather than human-based ones, which makes their applicability in humans a question. Although the cGAS-STING pathway is highly involved in the inflammatory mechanism of COPD, it is merely a fraction of the overall picture, necessitating further exploration of additional mechanisms. Moreover, inhibitors of the cGAS-STING signaling pathway have not been extensively employed as COPD therapeutic drugs, and the mechanisms by which certain drugs function as inhibitors remain unclear.

In this review, we discuss the inflammatory pathogenesis of COPD and provide an overview of the current understanding of the cGAS-STING signaling pathway as well as its potential as a COPD therapeutic target. The cGAS-STING pathway is activated in COPD, and its activation further exacerbates the development of COPD. Targeting the cGAS-STING signaling pathway is highly important for curing COPD, and many studies have suggested that small molecule inhibitors are effective controlling the development of COPD. However, whether COPD is related to cGAS and STING remains an area that requires further research. In conclusion, targeting the cGAS-STING signaling pathway provides a promising direction for COPD therapy and intervention.

Abbreviations

COPD	Chronic Obstructive Pulmonary Disease
cGAS	cyclic GMP-AMP Synthase
STING	Stimulator of Interferon Genes
CS	Cigarette Smoking
PRRs	Pattern Recognition Receptors
cgamp	cyclic GMP-AMP
ER	Endoplasmic Reticulum
GOLD	The Global Initiative of Obstructive Lung Disease
DAMPs	Damage-Associated Molecular Patterns
IL	Interleukin
TSLP	Thymic Stromal Lymphopoietin
ILC	Innate Lymphoid Cells
MCP	Monocyte Chemotactic Proteins
CCL	C-C motif chemokine ligand
dsDNA	double-stranded DNA
PAMPs	Pathogen Associated Molecular Patterns
DAMP	Damage Associated Molecular Pattern
TBK1	TANK-binding Kinase 1
IRF3	Interferon Regulatory Factor 3
ISGs	Immune Stimulating Genes
IFNs	Interferons
IKK	IkB Kinase
ERGIC	ER-Golgi Intermediate Compartment
Se Met	Selenomethionine
PPI	Protein – protein Interface
SASP	Senescence Associated Secretory Phenotype
TRQ	Tanreqing
PaO2	Partial Pressure of Oxygen
PaCO2	Pressure of Carbon Dioxide
MUC5A	Mucin 5AC
Ginseng	Panax Ginseng C.A Meyer
PPD	Protopanaxadiol
PPT	Protopanaxatriol
ROS	Reactive Oxygen Species

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Author contributions

J.R. provided the main direction and important guidance for this manuscript. K.L., F.W. and C.X. conceived the paper. K.L., Z.X., S.Z. and W.B. wrote the original draft and illustrated the figure for the manuscript. All authors approved the final manuscript.

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Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

Not applicable.

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Competing interests

The authors declare no competing interests.

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References

- Chen S, Kuhn M, Prettner K, Yu F, Yang T, Barnighausen T, Bloom DE, Wang C. The global economic burden of chronic obstructive pulmonary disease for 204 countries and territories in 2020-50: a health-augmented macroeconomic modelling study. LANCET GLOB HEALTH. 2023;11(8):e1183–93.
- Agusti A, Celli BR, Criner GJ, Halpin D, Anzueto A, Barnes P, Bourbeau J, Han MK, Martinez FJ, Montes DOM, et al. Global Initiative for Chronic Obstructive Lung Disease 2023 Report: GOLD Executive Summary. AM J RESP CRIT CARE. 2023;207(7):819–37.
- Fletcher C, Peto R. The natural history of chronic airflow obstruction. Br Med J. 1977;1(6077):1645–8.
- Civril F, Deimling T, de Oliveira MC, Ablasser A, Moldt M, Witte G, Hornung V, Hopfner KP. Structural mechanism of cytosolic DNA sensing by cGAS. Nature. 2013;498(7454):332–7.
- Ishikawa H, Barber GN. STING is an endoplasmic reticulum adaptor that facilitates innate immune signalling. Nature. 2008;455(7213):674–8.
- Gulen MF, Samson N, Keller A, Schwabenland M, Liu C, Gluck S, Thacker W, Favre L, Mangeat B, Kroese LJ, et al. cGAS-STING drives ageing-related inflammation and neurodegeneration. Nature. 2023;620(7973):374–80.
- Nascimento M, Gombault A, Lacerda-Queiroz N, Panek C, Savigny F, Sbeity M, Bourinet M, Le Bert M, Riteau N, Ryffel B, et al. Self-DNA release and STINGdependent sensing drives inflammation to cigarette smoke in mice. SCI REP-UK. 2019;9(1):14848.
- Benmerzoug S, Rose S, Bounab B, Gosset D, Duneau L, Chenuet P, Mollet L, Le Bert M, Lambers C, Geleff S, et al. STING-dependent sensing of self-DNA drives silica-induced lung inflammation. NAT COMMUN. 2018;9(1):5226.
- Wang X, Lu W, Xia X, Zhu Y, Ge C, Guo X, Zhang N, Chen H, Xu S. Selenomethionine mitigate PM2.5-induced cellular senescence in the lung via attenuating inflammatory response mediated by cGAS/STING/NF-kappaB pathway. ECOTOX ENVIRON SAFE. 2022;247(12):114266.
- Gluck S, Guey B, Gulen MF, Wolter K, Kang TW, Schmacke NA, Bridgeman A, Rehwinkel J, Zender L, Ablasser A. Innate immune sensing of cytosolic chromatin fragments through cGAS promotes senescence. NAT CELL BIOL. 2017;19(9):1061–70.
- Yang H, Wang H, Ren J, Chen Q, Chen ZJ. cGAS is essential for cellular senescence. P NATL ACAD SCI USA. 2017;114(23):E4612–20.
- Chilosi M, Carloni A, Rossi A, Poletti V. Premature lung aging and cellular senescence in the pathogenesis of idiopathic pulmonary fibrosis and COPD/ emphysema. TRANSL RES. 2013;162(3):156–73.
- Celli BR, Locantore N, Yates J, Tal-Singer R, Miller BE, Bakke P, Calverley P, Coxson H, Crim C, Edwards LD, et al. Inflammatory biomarkers improve clinical prediction of mortality in chronic obstructive pulmonary disease. AM J RESP CRIT CARE. 2012;185(10):1065–72.
- Singh D, Edwards L, Tal-Singer R, Rennard S. Sputum neutrophils as a biomarker in COPD: findings from the ECLIPSE study. RESP RES. 2010;11(1):77.
- Mendy A, Forno E, Niyonsenga T, Gasana J. Blood biomarkers as predictors of long-term mortality in COPD. CLIN RESPIR J. 2018;12(5):1891–9.

- Zhang MY, Jiang YX, Yang YC, Liu JY, Huo C, Ji XL, Qu YQ. Cigarette smoke extract induces pyroptosis in human bronchial epithelial cells through the ROS/NLRP3/caspase-1 pathway. LIFE SCI. 2021;269:119090.
- Shyam PSB, Chaya SK, Kumar VS, Mahendra M, Jayaraj BS, Lokesh KS, Ganguly K, Mahesh PA. Inflammatory Biomarkers Interleukin 1 Beta (IL-1beta) and Tumour Necrosis Factor Alpha (TNF-alpha) Are Differentially Elevated in Tobacco Smoke Associated COPD and Biomass Smoke Associated COPD. *TOXICS* 2021, 9(4).
- Benjamin JT, Plosa EJ, Sucre JM, van der Meer R, Dave S, Gutor S, Nichols DS, Gulleman PM, Jetter CS, Han W et al. Neutrophilic inflammation during lung development disrupts elastin assembly and predisposes adult mice to COPD. J CLIN INVEST 2021, 131(1).
- Genschmer KR, Russell DW, Lal C, Szul T, Bratcher PE, Noerager BD, Abdul RM, Xu X, Rezonzew G, Viera L, et al. Activated PMN exosomes: pathogenic entities causing Matrix Destruction and Disease in the lung. Cell. 2019;176(1–2):113–26.
- Mahor D, Kumari V, Vashisht K, Galgalekar R, Samarth RM, Mishra PK, Banerjee N, Dixit R, Saluja R, De S, et al. Elevated serum matrix metalloprotease (MMP-2) as a candidate biomarker for stable COPD. BMC PULM MED. 2020;20(1):302.
- Aida Y, Shibata Y, Abe S, Inoue S, Kimura T, Igarashi A, Yamauchi K, Nunomiya K, Kishi H, Nemoto T, et al. Inhibition of elastase-pulmonary emphysema in dominant-negative MafB transgenic mice. INT J BIOL SCI. 2014;10(8):882–94.
- Guyot N, Wartelle J, Malleret L, Todorov AA, Devouassoux G, Pacheco Y, Jenne DE, Belaaouaj A. Unopposed cathepsin G, neutrophil elastase, and proteinase 3 cause severe lung damage and emphysema. AM J PATHOL. 2014;184(8):2197–210.
- Guo Y, Ma L, Zhang F, Sun R, Li T. Neutrophil elastase ameliorates matrix metalloproteinase-9 to promote lipopolysaccharide-induced acute lung injury in mice 1. ACTA CIR BRAS. 2016;31(6):382–8.
- Arai N, Kondo M, Izumo T, Tamaoki J, Nagai A. Inhibition of neutrophil elastase-induced goblet cell metaplasia by tiotropium in mice. EUR RESPIR J. 2010;35(5):1164–71.
- Yun JH, Lamb A, Chase R, Singh D, Parker MM, Saferali A, Vestbo J, Tal-Singer R, Castaldi PJ, Silverman EK, et al. Blood eosinophil count thresholds and exacerbations in patients with chronic obstructive pulmonary disease. J ALLERGY CLIN IMMUN. 2018;141(6):2037–47.
- Eltboli O, Bafadhel M, Hollins F, Wright A, Hargadon B, Kulkarni N, Brightling C. COPD exacerbation severity and frequency is associated with impaired macrophage efferocytosis of eosinophils. BMC PULM MED. 2014;14:112.
- Ying S, Robinson DS, Meng Q, Rottman J, Kennedy R, Ringler DJ, Mackay CR, Daugherty BL, Springer MS, Durham SR, et al. Enhanced expression of eotaxin and CCR3 mRNA and protein in atopic asthma. Association with airway hyperresponsiveness and predominant co-localization of eotaxin mRNA to bronchial epithelial and endothelial cells. EUR J IMMUNOL. 1997;27(12):3507–16.
- Hirai H, Tanaka K, Yoshie O, Ogawa K, Kenmotsu K, Takamori Y, Ichimasa M, Sugamura K, Nakamura M, Takano S, et al. Prostaglandin D2 selectively induces chemotaxis in T helper type 2 cells, eosinophils, and basophils via seven-transmembrane receptor CRTH2. J EXP MED. 2001;193(2):255–61.
- 29. Davoine F, Lacy P. Eosinophil cytokines, chemokines, and growth factors: emerging roles in immunity. FRONT IMMUNOL. 2014;5:570.
- Bafadhel M, Peterson S, De Blas MA, Calverley PM, Rennard SI, Richter K, Fageras M. Predictors of exacerbation risk and response to budesonide in patients with chronic obstructive pulmonary disease: a post-hoc analysis of three randomised trials. LANCET RESP MED. 2018;6(2):117–26.
- Hoffmann A, Levchenko A, Scott ML, Baltimore D. The IkappaB-NF-kappaB signaling module: temporal control and selective gene activation. Science. 2002;298(5596):1241–5.
- 32. Lee UJ, Choung SR, Prakash KV, Lee EJ, Lee MY, Kim YJ, Han CW, Choi YC. Dual knockdown of p65 and p50 subunits of NF-kappaB by siRNA inhibits the induction of inflammatory cytokines and significantly enhance apoptosis in human primary synoviocytes treated with tumor necrosis factor-alpha. MOL BIOL REP. 2008;35(3):291–8.
- Zhuan B, Yu Y, Yang Z, Zhao X, Li P. Mechanisms of oxidative stress effects of the NADPH oxidase-ROS-NF-kappaB transduction pathway and VPO1 on patients with chronic obstructive pulmonary disease combined with pulmonary hypertension. EUR REV MED PHARMACO. 2017;21(15):3459–64.
- Xu H, Sun Q, Lu L, Luo F, Zhou L, Liu J, Cao L, Wang Q, Xue J, Yang Q, et al. MicroRNA-218 acts by repressing TNFR1-mediated activation of NF-kappaB, which is involved in MUC5AC hyper-production and inflammation in smoking-induced bronchiolitis of COPD. TOXICOL LETT. 2017;280:171–80.

- Kaur G, Batra S. Regulation of DNA methylation signatures on NF-kappaB and STAT3 pathway genes and TET activity in cigarette smoke extractchallenged cells/COPD exacerbation model in vitro. CELL BIOL TOXICOL. 2020;36(5):459–80.
- Brancho D, Tanaka N, Jaeschke A, Ventura JJ, Kelkar N, Tanaka Y, Kyuuma M, Takeshita T, Flavell RA, Davis RJ. Mechanism of p38 MAP kinase activation in vivo. GENE DEV. 2003;17(16):1969–78.
- Gaffey K, Reynolds S, Plumb J, Kaur M, Singh D. Increased phosphorylated p38 mitogen-activated protein kinase in COPD lungs. EUR RESPIR J. 2013;42(1):28–41.
- Knobloch J, Jungck D, Kronsbein J, Stoelben E, Ito K, Koch A. LABAs and p38MAPK inhibitors reverse the corticosteroid-insensitivity of IL-8 in Airway smooth muscle cells of COPD. J CLIN MED 2019, 8(12).
- Oh SY, Kim YH, Kang MK, Lee EJ, Kim DY, Oh H, Kim SI, Na W, Kang IJ, Kang YH. Aesculetin inhibits airway thickening and mucus Overproduction Induced by Urban Particulate Matter through blocking inflammation and oxidative stress involving TLR4 and EGFR. ANTIOXIDANTS-BASEL 2021, 10(3).
- Fang S, Crews AL, Chen W, Park J, Yin Q, Ren XR, Adler KB. MARCKS and HSP70 interactions regulate mucin secretion by human airway epithelial cells in vitro. AM J PHYSIOL-LUNG C. 2013;304(8):L511–8.
- Lai Y, Fois G, Flores JR, Tuvim MJ, Zhou Q, Yang K, Leitz J, Peters J, Zhang Y, Pfuetzner RA, et al. Inhibition of calcium-triggered secretion by hydrocarbonstapled peptides. Nature. 2022;603(7903):949–56.
- 42. Cloonan SM, Lam HC, Ryter SW, Choi AM. Ciliophagy: the consumption of cilia components by autophagy. AUTOPHAGY. 2014;10(3):532–4.
- Sun L, Wu J, Du F, Chen X, Chen ZJ. Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway. Science. 2013;339(6121):786–91.
- 44. Du M, Chen ZJ. DNA-induced liquid phase condensation of cGAS activates innate immune signaling. Science. 2018;361(6403):704–9.
- Kranzusch PJ, Lee AS, Berger JM, Doudna JA. Structure of human cGAS reveals a conserved family of second-messenger enzymes in innate immunity. CELL REP. 2013;3(5):1362–8.
- Diner EJ, Burdette DL, Wilson SC, Monroe KM, Kellenberger CA, Hyodo M, Hayakawa Y, Hammond MC, Vance RE. The innate immune DNA sensor cGAS produces a noncanonical cyclic dinucleotide that activates human STING. CELL REP. 2013;3(5):1355–61.
- Blander JM, Sander LE. Beyond pattern recognition: five immune checkpoints for scaling the microbial threat. NAT REV IMMUNOL. 2012;12(3):215–25.
- Ablasser A, Goldeck M, Cavlar T, Deimling T, Witte G, Rohl I, Hopfner KP, Ludwig J, Hornung V. cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates STING. Nature. 2013;498(7454):380–4.
- Gao P, Ascano M, Wu Y, Barchet W, Gaffney BL, Zillinger T, Serganov AA, Liu Y, Jones RA, Hartmann G, et al. Cyclic [G(2/5')pA(3/5')p] is the metazoan second messenger produced by DNA-activated cyclic GMP-AMP synthase. Cell. 2013;153(5):1094–107.
- Li XD, Wu J, Gao D, Wang H, Sun L, Chen ZJ. Pivotal roles of cGAS-cGAMP signaling in antiviral defense and immune adjuvant effects. Science. 2013;341(6152):1390–4.
- Shang G, Zhang C, Chen ZJ, Bai XC, Zhang X. Cryo-EM structures of STING reveal its mechanism of activation by cyclic GMP-AMP. Nature. 2019;567(7748):389–93.
- 52. Zhao B, Du F, Xu P, Shu C, Sankaran B, Bell SL, Liu M, Lei Y, Gao X, Fu X et al. A conserved PLPLRT/SD motif of STING mediates the recruitment and activation of TBK1. *NATURE* 2019, 569(7758):718–722.
- Zhang C, Shang G, Gui X, Zhang X, Bai XC, Chen ZJ. Structural basis of STING binding with and phosphorylation by TBK1. NATURE 2019, 567(7748):394–398.
- Liu S, Cai X, Wu J, Cong Q, Chen X, Li T, Du F, Ren J, Wu YT, Grishin NV, et al. Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3 activation. Science. 2015;347(6227):aaa2630.
- Gonugunta VK, Sakai T, Pokatayev V, Yang K, Wu J, Dobbs N, Yan N. Traffickingmediated STING degradation requires sorting to Acidified endolysosomes and can be targeted to Enhance Anti-tumor Response. CELL REP. 2017;21(11):3234–42.
- Fitzgerald KA, McWhirter SM, Faia KL, Rowe DC, Latz E, Golenbock DT, Coyle AJ, Liao SM, Maniatis T. IKKepsilon and TBK1 are essential components of the IRF3 signaling pathway. NAT IMMUNOL. 2003;4(5):491–6.
- Wu J, Sun L, Chen X, Du F, Shi H, Chen C, Chen ZJ. Cyclic GMP-AMP is an endogenous second messenger in innate immune signaling by cytosolic DNA. Science. 2013;339(6121):826–30.

- Gentili M, Lahaye X, Nadalin F, Nader G, Puig LE, Herve S, De Silva NS, Rookhuizen DC, Zueva E, Goudot C, et al. The N-Terminal Domain of cGAS Determines Preferential Association with centromeric DNA and Innate Immune activation in the Nucleus. CELL REP. 2019;26(9):2377–93.
- 60. Volkman HE, Cambier S, Gray EE, Stetson DB. Tight nuclear tethering of cGAS is essential for preventing autoreactivity. *ELIFE* 2019, 8.
- Jiang H, Xue X, Panda S, Kawale A, Hooy RM, Liang F, Sohn J, Sung P, Gekara NO. Chromatin-bound cGAS is an inhibitor of DNA repair and hence accelerates genome destabilization and cell death. EMBO J. 2019;38(21):e102718.
- Gao D, Li T, Li XD, Chen X, Li QZ, Wight-Carter M, Chen ZJ. Activation of cyclic GMP-AMP synthase by self-DNA causes autoimmune diseases. P NATL ACAD SCI USA. 2015;112(42):E5699–705.
- Kerur N, Fukuda S, Banerjee D, Kim Y, Fu D, Apicella I, Varshney A, Yasuma R, Fowler BJ, Baghdasaryan E, et al. cGAS drives noncanonical-inflammasome activation in age-related macular degeneration. NAT MED. 2018;24(1):50–61.
- Pouwels SD, Zijlstra GJ, van der Toorn M, Hesse L, Gras R, Ten HN, Krysko DV, Vandenabeele P, de Vries M, van Oosterhout AJ, et al. Cigarette smokeinduced necroptosis and DAMP release trigger neutrophilic airway inflammation in mice. AM J PHYSIOL-LUNG C. 2016;310(4):L377–86.
- 65. Sears CR. DNA repair as an emerging target for COPD-lung cancer overlap. RESPIR INVESTIG. 2019;57(2):111–21.
- You R, Lu W, Shan M, Berlin JM, Samuel EL, Marcano DC, Sun Z, Sikkema WK, Yuan X, Song L, et al. Nanoparticulate carbon black in cigarette smoke induces DNA cleavage and Th17-mediated emphysema. ELIFE. 2015;4:e9623.
- 67. Ahn J, Gutman D, Saijo S, Barber GN. STING manifests self DNA-dependent inflammatory disease. P NATL ACAD SCI USA. 2012;109(47):19386–91.
- 68. Zhu HF, Li Y. Small-molecule targets in Tumor Immunotherapy. NAT Prod BIOPROSP. 2018;8(4):297–301.
- An J, Woodward JJ, Sasaki T, Minie M, Elkon KB. Cutting edge: antimalarial drugs inhibit IFN-beta production through blockade of cyclic GMP-AMP synthase-DNA interaction. J IMMUNOL. 2015;194(9):4089–93.
- An J, Woodward JJ, Lai W, Minie M, Sun X, Tanaka L, Snyder JM, Sasaki T, Elkon KB. Inhibition of cyclic GMP-AMP synthase using a Novel Antimalarial Drug Derivative in Trex1-Deficient mice. ARTHRITIS RHEUMATOL. 2018;70(11):1807–19.
- Steinhagen F, Zillinger T, Peukert K, Fox M, Thudium M, Barchet W, Putensen C, Klinman D, Latz E, Bode C. Suppressive oligodeoxynucleotides containing TTAGGG motifs inhibit cGAS activation in human monocytes. EUR J IMMU-NOL. 2018;48(4):605–11.
- Wang M, Sooreshjani MA, Mikek C, Opoku-Temeng C, Sintim HO. Suramin potently inhibits cGAMP synthase, cGAS, in THP1 cells to modulate IFN-beta levels. FUTURE MED CHEM. 2018;10(11):1301–17.
- Vincent J, Adura C, Gao P, Luz A, Lama L, Asano Y, Okamoto R, Imaeda T, Aida J, Rothamel K, et al. Small molecule inhibition of cGAS reduces interferon expression in primary macrophages from autoimmune mice. NAT COMMUN. 2017;8(1):750.
- Zhao W, Xiong M, Yuan X, Li M, Sun H, Xu Y. In Silico Screening-based Discovery of Novel inhibitors of human cyclic GMP-AMP synthase: a Cross-validation Study of Molecular Docking and experimental testing. J CHEM INF MODEL. 2020;60(6):3265–76.
- Siu T, Altman MD, Baltus GA, Childers M, Ellis JM, Gunaydin H, Hatch H, Ho T, Jewell J, Lacey BM, et al. Discovery of a novel cGAMP competitive ligand of the inactive form of STING. ACS MED CHEM LETT. 2019;10(1):92–7.
- Li S, Hong Z, Wang Z, Li F, Mei J, Huang L, Lou X, Zhao S, Song L, Chen W, et al. The Cyclopeptide Astin C specifically inhibits the Innate Immune CDN Sensor STING. CELL REP. 2018;25(12):3405–21.
- Haag SM, Gulen MF, Reymond L, Gibelin A, Abrami L, Decout A, Heymann M, van der Goot FG, Turcatti G, Behrendt R, et al. Targeting STING with covalent small-molecule inhibitors. Nature. 2018;559(7713):269–73.
- Padilla-Salinas R, Sun L, Anderson R, Yang X, Zhang S, Chen ZJ, Yin H. Discovery of Small-Molecule Cyclic GMP-AMP synthase inhibitors. J ORG CHEM. 2020;85(3):1579–600.
- Huffman BJ, Chen S, Schwarz JL, Plata RE, Chin EN, Lairson LL, Houk KN, Shenvi RA. Electronic complementarity permits hindered butenolide heterodimerization and discovery of novel cGAS/STING pathway antagonists. NAT CHEM. 2020;12(3):310–7.
- 80. Hall J, Brault A, Vincent F, Weng S, Wang H, Dumlao D, Aulabaugh A, Aivazian D, Castro D, Chen M, et al. Discovery of PF-06928215 as a high affinity

inhibitor of cGAS enabled by a novel fluorescence polarization assay. PLoS ONE. 2017;12(9):e184843.

- Vinogradova EV, Zhang X, Remillard D, Lazar DC, Suciu RM, Wang Y, Bianco G, Yamashita Y, Crowley VM, Schafroth MA et al. An Activity-Guided Map of Electrophile-Cysteine Interactions in Primary Human T Cells. *CELL* 2020, 182(4):1009–1026.
- Gomez AM, Muino MA, Bendala EA, Maestro DLCG, Monge MD, Boixeda R, Ena J, Mella PC, Anton SJ, Lumbreras BC. Clinical characteristics and prognosis of COPD patients hospitalized with SARS-CoV-2. INT J CHRONIC OBSTR. 2020;15:3433–45.
- Zhou Y, Hou Y, Shen J, Huang Y, Martin W, Cheng F. Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2. CELL DISCOV. 2020;6:14.
- Huttner A, Kowalczyk A, Turjeman A, Babich T, Brossier C, Eliakim-Raz N, Kosiek K, Martinez DTB, Roux X, Shiber S, et al. Effect of 5-Day nitrofurantoin vs single-dose fosfomycin on clinical resolution of uncomplicated lower urinary tract infection in women: a Randomized Clinical Trial. JAMA-J AM MED ASSOC. 2018;319(17):1781–9.
- Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: an expanding universe. Cell. 2023;186(2):243–78.
- Tsuji T, Aoshiba K, Nagai A. Alveolar cell senescence in patients with pulmonary emphysema. AM J RESP CRIT CARE. 2006;174(8):886–93.
- Rutten EP, Gopal P, Wouters EF, Franssen FM, Hageman GJ, Vanfleteren LE, Spruit MA, Reynaert NL. Various mechanistic pathways representing the aging process are altered in COPD. Chest. 2016;149(1):53–61.
- Rivas M, Gupta G, Costanzo L, Ahmed H, Wyman AE, Geraghty P. Senescence: pathogenic driver in Chronic Obstructive Pulmonary Disease. MEDICINA-LITHUANIA 2022, 58(6).
- Tsuji T, Aoshiba K, Nagai A. Cigarette smoke induces senescence in alveolar epithelial cells. AM J RESP CELL MOL. 2004;31(6):643–9.
- Fujii S, Hara H, Araya J, Takasaka N, Kojima J, Ito S, Minagawa S, Yumino Y, Ishikawa T, Numata T, et al. Insufficient autophagy promotes bronchial epithelial cell senescence in chronic obstructive pulmonary disease. ONCO-IMMUNOLOGY. 2012;1(5):630–41.
- 91. Lan YY, Heather JM, Eisenhaure T, Garris CS, Lieb D, Raychowdhury R, Hacohen N. Extranuclear DNA accumulates in aged cells and contributes to senescence and inflammation. Aging Cell. 2019;18(2):e12901.
- 92. Liu W, Zhang X, Mao B, Jiang H. Systems pharmacology-based study of tanreqing injection in airway mucus hypersecretion. J ETHNOPHARMACOL. 2020;249:112425.
- Deng J, He Y, Sun G, Yang H, Wang L, Tao X, Chen W. Tanreqing injection protects against bleomycin-induced pulmonary fibrosis via inhibiting STINGmediated endoplasmic reticulum stress signaling pathway. J ETHNOPHAR-MACOL. 2023;305:116071.
- 94. Choi KT. Botanical characteristics, pharmacological effects and medicinal components of Korean Panax ginseng C a Meyer. ACTA PHARMACOL SIN. 2008;29(9):1109–18.
- Cho JY, Yoo ES, Baik KU, Park MH, Han BH. In vitro inhibitory effect of protopanaxadiol ginsenosides on tumor necrosis factor (TNF)-alpha production and its modulation by known TNF-alpha antagonists. PLANTA MED. 2001;67(3):213–8.
- Guan X, Yuan Y, Wang G, Zheng R, Zhang J, Dong B, Ran N, Hsu AC, Wang C, Wang F. Ginsenoside Rg3 ameliorates acute exacerbation of COPD by suppressing neutrophil migration. INT IMMUNOPHARMACOL. 2020;83:106449.
- Li H, Cui L, Liu Q, Dou S, Wang W, Xie M, Xu X, Zheng C, Li T, Huang S, et al. Ginsenoside Rb3 alleviates CSE-induced TROP2 upregulation through p38 MAPK and NF-kappaB pathways in basal cells. AM J RESP CELL MOL. 2021;64(6):747–59.
- Shergis JL, Di YM, Zhang AL, Vlahos R, Helliwell R, Ye JM, Xue CC. Therapeutic potential of Panax ginseng and ginsenosides in the treatment of chronic obstructive pulmonary disease. COMPLEMENT THER MED. 2014;22(5):944–53.
- Pang W, Lin S, Dai Q, Zhang H, Hu J. Antitussive activity of Pseudostellaria heterophylla (miq.) Pax extracts and improvement in lung function via adjustment of multi-cytokine levels. Molecules. 2011;16(4):3360–70.
- 100. Sun SC, Han R, Hou SS, Yi HQ, Chi SJ, Zhang AH. Juglanin alleviates bleomycin-induced lung injury by suppressing inflammation and fibrosis via targeting sting signaling. BIOMED PHARMACOTHER. 2020;127:110119.

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